



## INTERNATIONAL APPLICATION PUBLISHED UNDER THE PATENT COOPERATION TREATY (PCT)

<b>(51) International Patent Classification <sup>7</sup> :</b> <b>C12N 15/07, 15/01, 5/06</b>	<b>A1</b>	<b>(11) International Publication Number:</b> <b>WO 00/44893</b> <b>(43) International Publication Date:</b> 3 August 2000 (03.08.00)
<b>(21) International Application Number:</b> PCT/US00/01867 <b>(22) International Filing Date:</b> 27 January 2000 (27.01.00) <b>(30) Priority Data:</b> 60/117,718                      28 January 1999 (28.01.99)                      US <b>(71) Applicant (for all designated States except US):</b> PALMETTO RICHLAND MEMORIAL HOSPITAL [US/US]; 7 Rich- land Medical Park Drive, Columbia, SC 29203 (US). <b>(72) Inventor; and</b> <b>(75) Inventor/Applicant (for US only):</b> LAMB, Lawrence, S., Jr. [US/US]; 1621 Nursery Hill Road, Columbia, SC 29212 (US). <b>(74) Agents:</b> HARDAWAY, John, B., III et al.; Nexsen Pruet Jacobs & Pollard, LLP, P.O. Drawer 10648, Greenville, SC 29603-0648 (US).		<b>(81) Designated States:</b> AE, AL, AM, AT, AU, AZ, BA, BB, BG, BR, BY, CA, CH, CN, CR, CU, CZ, DE, DK, DM, EE, ES, FI, GB, GD, GE, GH, GM, HR, HU, ID, IL, IN, IS, JP, KE, KG, KP, KR, KZ, LC, LK, LR, LS, LT, LU, LV, MA, MD, MG, MK, MN, MW, MX, NO, NZ, PL, PT, RO, RU, SD, SE, SG, SI, SK, SL, TJ, TM, TR, TT, TZ, UA, UG, US, UZ, VN, YU, ZA, ZW, ARIPO patent (GH, GM, KE, LS, MW, SD, SL, SZ, TZ, UG, ZW), Eurasian patent (AM, AZ, BY, KG, KZ, MD, RU, TJ, TM), European patent (AT, BE, CH, CY, DE, DK, ES, FI, FR, GB, GR, IE, IT, LU, MC, NL, PT, SE), OAPI patent (BF, BJ, CF, CG, CI, CM, GA, GN, GW, ML, MR, NE, SN, TD, TG).  <b>Published</b> <i>With international search report.</i> <i>Before the expiration of the time limit for amending the</i> <i>claims and to be republished in the event of the receipt of</i> <i>amendments.</i>
<b>(54) Title:</b> IN VITRO ACTIVATED GAMMA DELTA LYMPHOCYTES  <b>(57) Abstract</b> <p>Patients who develop increased numbers of <math>\gamma\delta</math>+ cytotoxic T lymphocytes 2-6 months after allogeneic bone marrow transplantation are less likely to relapse than those who do not. The <math>\gamma\delta</math>+ T cells isolated from blood of patients with increased <math>\gamma\delta</math>+ T cells are CD3+CD4-CD8-CD57+, cytolytic to K562 cells, and express the V<math>\delta</math>1 T cell receptor phenotype. Similar <math>\gamma\delta</math>+ T cells can be generated <i>in vitro</i> by culture of donor mononuclear cells which are enriched for <math>\gamma\delta</math>+ T cells by immunomagnetic depletion of depleted of CD4+ and CD8+ cells. This <math>\gamma\delta</math>-enriched cell preparation was cultured on a combination of immobilized pan-<math>\delta</math> monoclonal antibody and irradiated recipient B cell leukemia. After four weeks, the cultures were almost exclusively V<math>\delta</math>1+CD3+CD4-CD8- cells that co-expressed activation-associated antigens CD69, CD25, and HLA-DR. Furthermore, they were cytolytic against the primary leukemia obtained from the recipient and lymphoblastic leukemia cell lines, yet had minimal cytotoxicity against normal donor-derived mononuclear cells or myeloid leukemia cell lines. These observations suggest that donor-derived cytotoxic <math>\gamma\delta</math>+ T cells can be generated <i>in vitro</i>, and may provide therapeutic potential for prevention of disease relapse.</p>		

**FOR THE PURPOSES OF INFORMATION ONLY**

Codes used to identify States party to the PCT on the front pages of pamphlets publishing international applications under the PCT.

AL	Albania	ES	Spain	LS	Lesotho	SI	Slovenia
AM	Armenia	FI	Finland	LT	Lithuania	SK	Slovakia
AT	Austria	FR	France	LU	Luxembourg	SN	Senegal
AU	Australia	GA	Gabon	LV	Latvia	SZ	Swaziland
AZ	Azerbaijan	GB	United Kingdom	MC	Monaco	TD	Chad
BA	Bosnia and Herzegovina	GE	Georgia	MD	Republic of Moldova	TG	Togo
BB	Barbados	GH	Ghana	MG	Madagascar	TJ	Tajikistan
BE	Belgium	GN	Guinea	MK	The former Yugoslav Republic of Macedonia	TM	Turkmenistan
BF	Burkina Faso	GR	Greece	ML	Mali	TR	Turkey
BG	Bulgaria	HU	Hungary	MN	Mongolia	TT	Trinidad and Tobago
BJ	Benin	IE	Ireland	MR	Mauritania	UA	Ukraine
BR	Brazil	IL	Israel	MW	Malawi	UG	Uganda
BY	Belarus	IS	Iceland	MX	Mexico	US	United States of America
CA	Canada	IT	Italy	NE	Niger	UZ	Uzbekistan
CF	Central African Republic	JP	Japan	NL	Netherlands	VN	Viet Nam
CG	Congo	KE	Kenya	NO	Norway	YU	Yugoslavia
CH	Switzerland	KG	Kyrgyzstan	NZ	New Zealand	ZW	Zimbabwe
CI	Côte d'Ivoire	KP	Democratic People's Republic of Korea	PL	Poland		
CM	Cameroon	KR	Republic of Korea	PT	Portugal		
CN	China	KZ	Kazakhstan	RO	Romania		
CU	Cuba	LC	Saint Lucia	RU	Russian Federation		
CZ	Czech Republic	LI	Liechtenstein	SD	Sudan		
DE	Germany	LK	Sri Lanka	SE	Sweden		
DK	Denmark	LR	Liberia	SG	Singapore		
EE	Estonia						

## IN VITRO ACTIVATED GAMMA DELTA LYMPHOCYTES

### BACKGROUND OF THE INVENTION

Allogeneic bone marrow transplantation (BMT) provides a potentially curative treatment for leukemias that are refractory to conventional therapy. In addition to providing hematopoietic rescue from myeloablative therapy, BMT offers an adoptive immunotherapy effect (graft-versus-leukemia-GvL) that can be beneficial in the elimination of residual leukemia. This was initially shown in cases where T cell depletion (TCD) has been used to prevent graft-versus-host disease (GvHD) but also experienced an increase in disease relapse (1). Indeed, relapse rates in high-risk patients (long-standing recurrent disease or relapse at the time of BMT) can be as high as 70% (2). Therefore, further improvement in disease-free survival is likely to depend on the antileukemic effectiveness of the transplant, i.e. maximizing the GvL effect.

Most experimental evidence suggests that GvL effectors are predominantly T cells that can either recognize allospecific molecules expressed on both normal and neoplastic hematopoietic cells or recognize cell surface molecules that are either unique to or preferentially expressed by the leukemia (3-7). Identification of specific cell populations that are important antileukemic effectors is an essential first step to successful GvL graft engineering and cellular immunotherapy.

Although several studies have suggested that  $\gamma\delta$ + T cells may not be important primary effectors of GvHD (8-12), few have addressed the GvL potential of  $\gamma\delta$ + T cells. Esslin (13) noted that *in vitro* activated  $\gamma\delta$ + T cells can mediate broadly-based non-MHC restricted cytolytic activity to selected human tumor cell lines. Others have shown that  $\gamma\delta$ + T cells can recognize unprocessed peptides, some of which are preferentially expressed on tumor cells (14-18). Finally, one report has shown cytotoxic anti-leukemic activity in a patient against B cell ALL by  $\gamma\delta$ + T cells expressing the V $\delta$ 1 form of the T cell receptor (19). Taken together, these findings support a potential antileukemic role for  $\gamma\delta$ + T cells.

Published data describing a series of 10 leukemia patients who developed an increased proportion of circulating CD3+CD4-CD8-V $\delta$ 1+ $\gamma\delta$ + T cells between 60

and 270 days post-BMT from a partially mismatched related donor (PMRD) which continued for up to two years. Eight of these patients are surviving and remain free of disease, as compared to a DFS probability of 31% at 2.5 years among 100-day survivors with a normal number of  $\gamma\delta$ + T cells (20). In addition, it has  
5 been recently shown that enrichment of the graft with  $\gamma\delta$ + T cells may have contributed to the later development of increased  $\gamma\delta$ + T cells (21). Regardless of the TCD protocol used, however, patients who developed increased  $\gamma\delta$ + T cells showed the same cell phenotype and cytolytic function as well as a decreased incidence of relapse.

10       **Allogeneic Bone Marrow Transplantation and Graft-Host Interactions:**  
High-dose chemo/radiotherapy followed by bone marrow rescue provides a potentially curative treatment for a variety of leukemias and solid tumors that are refractory to conventional therapy. An alloreactive response, mediated by donor immunocompetent cells in the graft and directed against normal cells and tissues  
15 in the recipient can result in the development of graft-versus-host disease (GvHD). GvHD can occur in up to 50% of patients receiving unmodified, HLA-identical marrow, indicating that minor histocompatibility differences, not detected by conventional HLA matching techniques, can initiate this reaction (22,23). For the majority of patients (approximately 70%) who do not have matched sibling donor  
20 (MSD) alternative donors may be used but the risk of acute GvHD is increased due to differences in major as well as minor histocompatibility antigens (1). The same alloreactive response, however, can be beneficial in the elimination of residual leukemia through an adoptive immunotherapy mechanism known as the graft-versus-leukemia (GvL effect).

25       **Allogeneic BMT and the use of Alternative Donors:** In most instances, the ideal bone marrow donor is the HLA-identical sibling. Alternative donors include the HLA-phenotypically matched unrelated donor (MUD), a partially mismatched related donor (PMRD) or a cord blood donor (CBD), who can be a phenotypically matched or mismatched related or unrelated donor (1).

30       **Graft engineering, T cell depletion, and graft-host interactions:** Initial attempts to use non-manipulated marrow from MUDs and PMRDs have resulted

in severe or fatal GvHD (24,25). This stimulated the development of methods to remove the suspected mediators of GvHD (T lymphocytes) from the marrow *ex vivo* prior to infusion (26). Results from transplants in which patients received marrow that was highly depleted of T cells (pan-T cell depletion) were initially  
5 promising, in that GvHD was significantly reduced; however, this was accompanied by an increase in graft failure (27,28), suggesting that donor T cells may eliminate the ability of residual recipient T cells to reject the graft.

Animal studies of PMRD transplants have indicated that both CD4 and CD8-positive cells are capable of mediating lethal GvHD (29). Initial human  
10 studies have therefore used *ex vivo* pan-T cell depletion to engineer these grafts. This has either been achieved by agglutination with soybean lectin and rosetting the residual T cells with sheep red blood cells, or by use of T cell-directed MAbs, e.g. anti-CD2, CD3, CD5, in combination with panning or complement to eliminate antibody-sensitized cells (26). In a study comparing 470 PMRD reduced the risk  
15 of acute GvHD, but increased the risk of graft failure, and there was no overall improvement in leukemia-free survival (30). Therefore, aggressive *ex vivo* pan-TCD was felt not to be optimal in facilitating PMRD BMT, and subsequent studies have explored the use of a modified pan-T cell depletion that leaves more T cells in the graft. Another option is the use of a more selective or targeted type of TCD  
20 often combined with post-transplant immune suppression (11-13).

When T cell depletion (TCD) has been used in matched sibling transplantation, a further concern has been an increase in disease relapse seen particularly in patients with CML (33). This apparent disruption in the graft-versus-leukemia (GvL) effect has discouraged investigators from using TCD other than  
25 when MHC-nonidentical grafts are used. We have, however, shown that the use of sequential immunomodulation of the patient and T cell depletion of up to 3Ag PMRD grafts can result in stable and sustained engraftment in >95% of recipients with a low incidence of acute and chronic GvHD (32). Relapse rates in high-risk patients (long-standing recurrent disease or relapse at the time of BMT) can be as  
30 high as 70% (2). This indicates that even though it is possible to cross major histocompatibility barriers with successful engraftment and a low incidence of GvHD, further improvement in disease-free survival will depend on the

antileukemic effectiveness of the transplant. While this might be accomplished by performing the transplant earlier in the disease course, many patients will not be referred for allogeneic BMT until they have demonstrated resistance to conventional-dose therap. Thus, enhancement of the GvL effect may be an essential component of the curative potential of allogeneic BMT.

**Biology of the GvL Effect:** The GvL reaction is through to be most effective in chronic phase CML (34,35), although there is also evidence for a GvL effect in the acute leukemias (36). It is generally thought that T lymphocytes recognize and eliminate residual leukemia through both MHC restricted and non-restricted pathways (37). Targets for GvL may include minor and/or major mismatched histocompatibility antigens and/or leukemia-specific antigens (38,39). Every allogeneic BMT patient potentially could benefit from the alloreactive response, although the extent of this benefit varies depending on whether the leukemia expresses allogeneic antigens to a degree that triggers recognition and killing. It is known that patients who suffer from acute and chronic GvHD post-BMT often have a reduced rate of leukemic relapse (3,4), possibly due to more intense alloreactivity against residual host-derived leukemic cells. Many investigators have shown evidence that GvHD and GvL effects can be separated to some degree (5,7,40,41), although a system for engineering a GvL effect in total absence of GvHD has not been reliably demonstrated.

T cell recognition of leukemia-associated antigens is also through to be a potentially important means by which immunocompetent cells may recognize and eliminate residual leukemia. It is known that leukemia-reactive clones can be generated (15). Specific targets for leukemia-reactive clones remain the topic of intense investigation, and some potential leukemia-associated antigens have been identified (3,16-19) and are discussed below. The ability to identify and stimulate a GvL effect via either or both of these mechanisms may be of therapeutic importance in reducing the risk of relapse in patients who have received TCD grafts.

**$\gamma\delta$ + T lymphocytes:** Five to ten percent of T cells in normal peripheral blood bear the  $\gamma\delta$  receptor (42), although this number may be slightly higher in Asians and Blacks. Recent observations suggest that  $\gamma\delta$ + T cells play a

substantially different role in the immune system than that of  $\alpha\beta^+$  T cells. One of the most obvious differences is that most  $\gamma\delta^+$  T cells usually do not co-express CD4 or CD8, and therefore may develop normally in the absence of MHC class II molecules (43) since positive selection may not be required. Similarly, it is difficult to elicit a response of  $\gamma\delta^+$  T cells against allogeneic MHC class I or II antigens, and when it has been possible to obtain  $\gamma\delta^+$  T cell clones against peptide antigens, recognition of these peptides is usually not restricted by classical MHC molecules (44). In addition,  $\gamma\delta^+$  T cells tend to recognize intact rather than processed polypeptide (44).

While the requirements for activation of human  $\gamma\delta^+$  T cells are still poorly understood, it is clear that they are different from those of  $\alpha\beta^+$  T cells.  $\gamma\delta$  T cells do not require presentation of antigens in the context of the MHC Class I or Class II molecules for activation (45), however, they probably require CD28-mediated co-stimulation, and, following activation, show autocrine IL-2 production (46).

They can also be activated by anti-CD2 antibodies (47).  $\gamma\delta^+$  T cells which express CD25 have also been shown to adhere to fibronectin-coated plates via the VLA-4 receptor with subsequent expansion, and cross linking of VLA-4 and VLA-5 receptors result in co-stimulated expansion induced by an anti pan- $\delta$  monoclonal antibody (48). Recent evidence has also suggested that certain subtypes of  $\gamma\delta^+$  T cells, predominantly the  $\gamma\delta^+$  CD8 $\alpha\alpha^+$  homodimer population, may be resistant to Cyclosporin A (49).

**Potential role of TCR- $\gamma\delta^+$  T lymphocytes in allogeneic BMT:** While activation mechanisms for  $\gamma\delta^+$  T cells are just being elucidated, even less is known about the role of these cells in graft-host interactions. Ellison (50) reported an increase in peripheral  $\gamma\delta^+$  T cells in murine studies of acute GvHD following allogeneic non-TCD BMT (50). In that study, depletion of  $\gamma\delta^+$  T cells resulted in a significant decrease in GvHD-related mortality. Blazar (51) also has shown that murine  $\gamma\delta^+$  T cells can play a role in rejection, alloengraftment, and GvHD through recognition of the "nonclassical" MHC class Ib antigens.

Studies in humans have to this point been in conflict with murine studies. Norton (8) did not find  $\gamma\delta^+$  T cells to be effectors of epidermal damage in cutaneous GvHD. Viale (9) did note an increase in the ratio of V $\delta$ 1;V $\delta$ 2 cells in

patients with acute GvHD but the significance of this finding remained undetermined. Tsuji (11) showed that although  $\gamma\delta$ + T cells cannot produce GvHD on their own, host  $\gamma\delta$ + T cells were recruited into donor  $\alpha\beta$ + lesions where they were activated and induced to proliferate. Transitory increases in the ratio of CD4<sup>-</sup>CD8<sup>-</sup>  $\gamma\delta$ + T cells have been reported during the first four weeks post-BMT in patients treated by GM-CSF, but the cells return to normal levels within eight weeks post-BMT (10). In addition, increased  $\gamma\delta$ + T cells have been found in one (study to be associated with viral and fungal infections during the first year following TCD BMT in patients receiving either PMRD or MUD grafts (12). In the same study, increases in  $\gamma\delta$ + T cells were not found to be associated with GvHD.

The potential for a possible anti-tumor role for  $\gamma\delta$ + T cells was established by Esslin (13), who noted that *in vitro* activated peripheral blood  $\gamma\delta$ + T cells possess cytolytic activity to selected human tumor cell lines when compared to similarly activated  $\alpha\beta$ + T cells. This reactivity was not MHC restricted, but was dependent on interaction with LFA-1b/ICAM1 rather than the  $\gamma\delta$  receptor. These cells predominantly expressed the V $\gamma$ 9/V $\delta$ 2 form of the T cell receptor. Proliferate responses of both  $\alpha\beta$ + and  $\gamma\delta$ + T cells, however, were inhibited by MAbs to anti-HLA-A, -B, and -C. These findings suggest that  $\gamma\delta$ + T cells activated through the TCR have an advantage in non-MHC restricted cytotoxicity which may correlate with a GvL response. It is known that  $\gamma\delta$ + T cells respond to heat shock proteins (16-18), some of which may be expressed by lymphomas. Human alloreactive  $\gamma\delta$ + T cells have also been generated which recognize TCT.1 (Blast-1/CD48), an antigen broadly distributed on hematopoietic cells (52). These  $\gamma\delta$ + T cells preferentially expressed the V $\gamma$ 3/V $\delta$ 1 form of the T cell receptor. V $\delta$ 1+ cell activation has also been reported in response to EBV-transformed B cells (14,53), EBV-infected Burkitt lymphoma cells (53), and Daudi lymphoma cells (54). In addition, one recent report has shown cytotoxic anti-leukemic activity in a patient against B cell ALL by  $\gamma\delta$ + T cells expressing the V $\delta$ 1 form of the T cell receptor (19).

We have been able to expand *in vitro* donor-derived  $\gamma\delta$  T cells which have a striking resemblance to those seen in the patients described above. Donor mononuclear cells were depleted of CD4+/CD8+ T cells, and expanded on a combination of immobilized pan- $\delta$  monoclonal antibody and irradiated recipient B



cell leukemia. After initial culture and re-stimulation, the cultures expanded rapidly and contained almost exclusively  $V\delta 1+$   $\gamma\delta+$  T cells which expressed CD3, CD25, and CD69, but were CD4- and CD8- which are cytolytic to recipient leukemia and K562 cells but are minimally cytolytic to self MNC and third party leukemia. These observations suggest that donor-derived  $\gamma\delta+$  T cells can be generated *in vitro*, thus providing a potential mechanism for cellular immunotherapy of leukemia.

### BRIEF DESCRIPTION OF THE FIGURES

Fig. 1 shows the expansion of donor  $\gamma\delta+$  T cells in culture.

Fig. 2 shows the phenotypic analysis of proliferating  $\gamma\delta+$  T cells from cultures on pan- $\delta$  MAb with blasts.

Fig. 3 shows the phenotypic analysis of proliferating  $\gamma\delta+$  T cells from cultures on pan- $\delta$  MAb without blasts.

Fig. 4 shows the phenotype of  $\gamma\delta+$  T cells from Patient #1.

Fig. 5 shows the flow cytometric binding assay depicting the binding of activated donor  $\gamma\delta+$  T cells to recipient leukemic CD19+ blasts.

Fig. 6 shows the cytotoxicity of donor  $\gamma\delta+$  T cells.

Fig. 7 shows the cytotoxicity of expanded  $\gamma\delta+$  T cells against various cell lines.

Fig. 8 shows the cytotoxic effects of expanded  $\gamma\delta+$  T cells against other cell lines.

Fig. 9 shows the mRNA and surface expression of  $V\delta$  subtypes.

### DESCRIPTION OF THE PREFERRED EMBODIMENTS

#### EXPERIMENTAL PROTOCOLS

Donor/recipient pairs: Three patients who presented for BMT with relapsed acute lymphoblastic leukemia or induction failure and their HLA-partially mismatched related donors were enrolled in this study.

Cell preparation: For recipients, sufficient blood was drawn to obtain a minimum of  $2.5 \times 10^7$  leukemic cells, but less than 50ml, prior to the start of pre-BMT conditioning therapy. Leukemic cells from the recipient were separated from normal mononuclear cells (MNC) using density gradient centrifugation on Percoll using 30-40% gradient. If necessary, further purification was accomplished by immunomagnetic

depletion of normal T and NK cells. Purity of normal and blast monolayers were evaluated by flow cytometry using MAbs previously found to be diagnostic for the patient's leukemia. The cells were then cryopreserved at a concentration of  $20 \times 10^6/\text{ml}$  in AIM-5 medium (Gibco) with 15% fetal bovine serum (Gibco) and 10% DMSO and stored in liquid nitrogen until donor selection was complete. Up to 50 ml of peripheral blood was then obtained from the corresponding partially mismatched related donor. These donor-derived  $\gamma\delta^+$  T cells were purified in the MNC layer by negative selection using CD4+ and CD8+ immunomagnetic microspheres (Dynal) at a ratio of 5 microspheres:cell. Removal of CD4+ and CD8+ cells from peripheral blood effectively depleted >95% of  $\alpha\beta^+$  T cells.

The number of  $\gamma\delta^+$  T cells in the preparation and the effectiveness of the  $\alpha\beta^+$  T cell depletion was monitored by flow cytometry as described below using fluorochrome-conjugated antibodies to TCR- $\alpha\beta$ , TCR- $\gamma\delta$ , CD4, CD8, and CD3 (Becton-Dickinson Immunocytometry Systems-BDIS; San Jose, CA).

Culture and activation of  $\gamma\delta^+$  T cells: Cytotoxic  $\gamma\delta^+$  T cells were generated from donor-recipient pairs as follows: Tissue culture-treated 24 well plates were coated with  $10\mu\text{g}$  TCR- $\delta 1$  pan- $\delta$  monoclonal antibody (Endogen; Woburn, MA) in  $300\mu\text{g}$  PBS for 24h at  $4^\circ\text{C}$  to facilitate initial activation and expansion of  $\gamma\delta^+$  T cells as described by Esslin (13). Irradiated (50Gy) primary leukemic blasts that were obtained and cryopreserved prior to BMT were thawed, washed x 3, and re-suspended in AIM-5 Media with 15% FBS and 25IU of IL-2 at a concentration of  $1.0 \times 10^6$  cells/ml. Aliquots of 1ml of this suspension were plated on the coated wells. Following immunomagnetic depletion of CD4+ and CD8+ cells as described above, remaining donor-derived MNC were adjusted to a concentration of  $1.0 \times 10^6$  cells/ml, and aliquots of 1 ml were added to the previously plated recipient blasts. Control wells consisted of CD4+CD8+ depleted MNC plated on TCR- $\delta 1$  monoclonal antibody in the absence of blasts or blasts in the absence of monoclonal antibody. The cultures were examined daily for characteristic morphology of proliferating clusters. Media was refreshed twice weekly or as necessary dependent on the robustness of proliferation as determined by microscopic examination and the phenol red pH indicator in the media. After two weeks in culture, cells were photographed, and subcultured 1:2 or 1:4 as necessary onto a freshly coated plate. The  $\gamma\delta^+$  T cell + blast wells were re-stimulated with freshly thawed blasts at the same concentration used previously and assayed at this time and weekly thereafter for phenotype, V $\delta$  subtype, and absolute cell number. At week four, fold expansion was calculated and harvesting was

begun for phenotypic, molecular, and functional assays described below and for cryopreservation and storage as described above for future study. These assays were performed at 4 – 6 weeks of culture. The concentration of  $\gamma\delta$ + T cells measured on a biweekly basis determined the degree of  $\gamma\delta$ + T cell stimulation for each culture condition.

5 When necessary, proliferating cells were transferred onto pan- $\delta$  MAb-coated tissue cultured flasks (Becton Dickinson) and cultures were maintained for up to twelve weeks, at which time no further proliferation was observed.

Flow cytometry: Expanded/activated  $\gamma\delta$ + T cells were analyzed by four color flow cytometry for expression of CD45, CD3, CD4, CD8, CD19, CD56, CD25, HLA-DR, 10 CD69 (Becton Dickinson Immunocytometry Systems; San Jose, CA-BDIS), and V $\delta$ 1 (Endogen, Woburn, MA), TCR- $\gamma\delta$ , CD57, CD94, and V $\delta$ 1-V $\delta$ 3 (Coulter Immunotech; Miami, FL) using monoclonal antibodies conjugated with fluorescein isothiocyanate (FITC), phycoerythrin (PE), peridinin chlorophyll protein (PerCP), or allophycocyanan (APC). Recipient primary B cell leukemias were analyzed for 15 expression of CD19, CD10, CD45, CD7, CD20, CD23, sIgG $\kappa$ , sIgG $\lambda$ , HLA-ABC, and HLA-DR (all from BDIS). At least 50,000 ungated events were collected in a list mode file and cell subpopulations in the lymphocyte CD45/side scatter gate and CD3/side scatter gate are quantitated and expressed as a percentage of the total lymphocyte population. Analysis was performed on a FACS Calibur flow 20 cytometer using CellQuest software (BDIS).

Flow cytometric binding assays. Binding of donor  $\gamma\delta$ + T cells to specific targets was examined by flow cytometry. Donor  $\gamma\delta$ + T cells were incubated in AIM-5 Media with 15% FBS for 30 minutes at 37°C, centrifuged, and resuspended in phosphate-buffered saline. The cell suspension was labeled with one MAb specific for 25 the leukemia but not expressed on  $\gamma\delta$ + T cells (CD19) and anti-TCR  $\gamma\delta$ , which is not expressed on the leukemia. The cell preparation was incubated at 4°C for 30 min, washed x 3, and analyzed by flow cytometry as detailed above. Clusters which were positive for both CD19 and  $\gamma\delta$  were then examined for forward (FSC) and side scatter (SSC) to determine if the represented multi-cell clusters. Dual-positive cells with 30 increased FSC and SSC were scored as bound blast/ $\gamma\delta$ + T cell clusters. Controls consisted of cultures of resting and activated donor  $\gamma\delta$ + T cells co-cultured K562 cells.

K562 cells are autofluorescent, so labeling with a fluoro-chrome was unnecessary. Resting  $\gamma\delta$ + T cells do not bind K562 while activated  $\gamma\delta$ + T cells do.

Cytotoxicity assays: Third-party mononuclear cells, K562 erythroleukemia cells, and recipient primary leukemia were used as targets. Aliquots of target cells were labeled overnight with 3,3'-dioctadecyloxycarbocyanine (DiOC<sub>18</sub>) (Molecular Probes, Eugene, OR). The cells were then washed in phosphate buffered saline (PBS) and resuspended in RPMI-1640 with 10% fetal bovine serum (FBS) at a concentration of  $2 \times 10^4$  cells/ml. Control MNC and expanded  $\gamma\delta$ + T cells were suspended in RPMI-1640 and diluted to yield E:T ratios of 40:1-2.5:1 and added to the target cells. Aliquots of 130 $\mu$ l counterstaining solution consisting of propidium iodine (PI) and PBS (Molecular Probes) were then added to the cell mixtures. The tubes were pelleted by centrifugation at 1000 x g for 30 sec and then incubated for 4 hours. Following incubation, the tubes were acquired in a FACS Calibur flow cytometer (BDIS) and analyzed for green fluorescence (DiOC<sub>18</sub>-560nm) and red fluorescence (PI-630nm). Analysis on a two parameter histogram allows separation of live target cells (DiOC<sub>18</sub>+PI-) and membrane-compromised targets are (DiOC<sub>18</sub>+PI+) from which % cytotoxicity was calculated.

$\gamma\delta$  T cell Receptor Characterization: The clonal heterogeneity of  $\gamma\delta$ + T cells determined by flow cytometry was further evaluated using molecular approaches to assess  $\gamma\delta$  TCR variable gene expression using peripheral blood mononuclear cells (PBMC) collected from the BMT donors and the expanded  $\gamma\delta$ + T cells from derived from culture on the pan- $\delta$  MAb and co-culture with the recipient ALL. Total RNA was extracted from MNC or cultured cells by the acid-phenol guanidinium thiocyanate method (55) and reverse transcribed according to the GeneAmp RNA PCR protocol (Perkin-Elmer Cetus, Norwalk, CT). The cDNA product served as template for PCR amplifications utilizing  $\gamma\delta$  TCR gene family-specific primers according to established methods (56). PCR amplification products were analyzed by agarose gel electrophoresis in order to determine the number and identity of  $\gamma\delta$  TCR V gene families expressed in each sample. This analysis was facilitated by DNA blot hybridization with corresponding TCR C $\gamma$  - or C $\delta$  -horseradish peroxidase (HRP) conjugated oligonucleotide probes followed by

chemiluminescent detection (23). Amplified products were resolved on 4% sequencing gels and detected, due to the incorporation of fluorescent primers during amplification, using the Hitachi FMBIO-100 Fluorescent Imager or the ABI 377 (Perkin-Elmer) automated sequencer using Genescan™ software. This method (known as TCR spectratyping) provides a more refined assessment of  $\gamma\delta$  TCR clonal diversity in the specimens.

## **RESULTS**

*Immobilized pan- $\delta$  MAb alone and with and leukemic blasts stimulate  $\gamma\delta$  T cells.*

As shown in Figure 1,  $\gamma\delta$  T cells strongly proliferated in response to immobilized pan- $\delta$  MAb alone and a combination of immobilized pan- $\delta$  MAb and blasts.

Leukemic blasts alone did not support sustained proliferation of  $\gamma\delta$  T cells. It should be noted, however, that in one experiment  $\gamma\delta$  T cell proliferation occurred later in the culture than in the other two experiments.

*Immunophenotypic analysis of proliferating  $\gamma\delta$  T cell cultures.* Phenotypic

analysis revealed that proliferating  $\gamma\delta$  T cells from cultures on pan- $\delta$  MAb with blasts preferentially expressed V $\delta$ 1 (Figure 2) while  $\gamma\delta$  T cells proliferating on pan- $\delta$  MAb without blasts preferentially expressed V $\delta$ 2 (Figure 3). The  $\gamma\delta$  T cell cultures were predominantly CD3+CD4-CD8- and expressed activation-associated antigens CD69, CD25, and HLA-DR regardless of culture conditions (Figure 4).

*Functional analysis of  $\gamma\delta$  T cell cultures.* Cultured donor-derived  $\gamma\delta$  T cells from both culture methods were tested for their ability to bind and to lyse primary leukemia from the corresponding BMT recipient. Figure 5 shows that indeed donor  $\gamma\delta$  T cells will bind recipient leukemia. Donor  $\gamma\delta$  T cells were highly cytotoxic to recipient leukemia as well as the NK sensitive target cell line K562 (Figure 6). In one experiment, mild nonspecific cytotoxicity was seen against third party MNC. Different lytic profiles were seen which correlated with culture method and predominant V $\delta$  gene usage (Figures 7 & 8). V $\delta$ 1+ cells cultured on immobilized pan- $\delta$  MAb and recipient blasts lysed primary ALL from the recipient and K562 cells as well as lymphoid cell lines, but had essentially no activity

against myeloid cell lines. In contrast, V $\delta$ 2 clones from cultures expanded on pan- $\delta$  MAb alone showed cytotoxic activity against all targets:

*TCR repertoire analysis of  $\gamma\delta$ + T cells.* Polyclonal  $\gamma\delta$ + T cells from the healthy BMT donors expressed mRNA predominantly for V $\delta$ 2 followed by V $\delta$ 1 and the V $\delta$ 3 (Figure 9). Occasionally mRNA for V $\delta$ 4 and V $\delta$ 5 was seen. Examination of the V $\delta$  repertoire of  $\gamma\delta$  cells cultured on pan- $\delta$  MAb alone was essentially unchanged from the peripheral blood V $\delta$  repertoire. In contrast,  $\gamma\delta$ + T cells cultured on pan- $\delta$  MAb and blasts showed preferential expression of V $\delta$ 1, followed by V $\delta$ 2 and V $\delta$ 3. High resolution analysis of these PCR products revealed.

It will be apparent to those of ordinary skill in the art that many modifications and substitutions can be made without departing from the spirit and the scope of the present invention.

#### REFERENCES

1. O'Reilly RJ, Hansen JA, Kurtzberg J, Henslee-Downey PJ, Martelli M, Aversa F., Allogeneic marrow transplantation: approaches for the patient lacking a donor. In Schechter, G.P., and McArthur, J.R. (eds), *Hematology 1996: Education Program for the American Society of Hematology*, 132-46.
2. Henslee-Downey PJ, Abhyankar SH, Parrish RS, Pati AR, Godder KT, Neglia WJ, Goon-Johnson KS, Geier SS, Lee CG, Gee AP. Use of partially mismatched related donors extends access to allogeneic marrow transplant. *Blood* 89 (10):3864, 1997.
3. Horowitz, M.M., Gale, R.P., Sondel, P.M., Goldman, J.M. et al. Graft versus leukemia reactions after bone marrow transplantation. *Blood* 75, 555, 1990.
4. Henslee, P.J., Thompson, J.S., Romond, E.H., Doukas, M.A. et al. T cell depletion of HLA and haploidentical marrow reduces graft-versus-host disease but it may impair a graft-versus-leukemia effect. *Transpln. Proc.* 19, 2701, 1987
5. Sykes, M., Romick, M.L., Sachs, D.H. Interleukin 2 prevents graft-versus-host disease while preserving the graft-versus-leukemia effect of allogeneic T cells. *Proc. Nat. Acad. Sci.* 1990; 87: 5633

6. Truitt, R.L., Atasoylu, A.A. Contribution of CD4+ and CD8+ T cells to graft-versus-host disease and the graft-versus-leukemia reactivity after transplantation of MHC compatible bone marrow *Bone Marrow Transplantation* 1991; 8: 51.
- 5 7. Weiss, L., Lubin, I., Factorowich, I., Lapidot, Z., Reich, S., Reisner, Y., Slavin, S. Effective the graft-versus-leukemia effects independent of graft-versus-host disease after T cell depleted allogeneic bone marrow transplantation in a murine model of B cell leukemia/lymphoma. *J. Immunol.* 1994; 153: 2562.
- 10 8. Norton J, Al-Saffar N, Sloane JP. An immunohistological study of lymphocytes in human cutaneous graft-versus-host disease. *Bone Marrow Transplantation* 7:205, 1991.
9. Viale M, Ferrini S, Bacigalupo A. TCR positive lymphocytes after allogeneic bone marrow transplantation. *Bone Marrow Transplantation* 10:249, 1992.
- 15 10. Yabe M, Yabe H, Hattori K, Hinorhars T, Morimoto T, Kato S, Kusonoki A. Transition of T-cell receptor gamma/delta expressing double negative (CD4/CD8) lymphocytes after allogeneic bone marrow transplantation. *Bone Marrow Transplantation* 14:741, 1994.
- 20 11. Tsuji S, Char D, Bucy RP, Simonsen M, Chen C, Cooper MD. + T-cells are secondary participants in acute graft-versus-host reactions initiated by CD4+ T-cells. *European Journal of Immunology* 26: 420, 1996.
- 25 12. Cela ME, Holliday MS, Rooney CM, Richardson S, Alexander B, Krance RA, Brenner MK, Heslop HE. +T-lymphocyte regeneration after T-lymphocyte-depleted bone marrow transplantation from mismatched family members or matched unrelated donors. *Bone Marrow Transplantation* 17:243, 1996.
13. Esslin A, Formby B. Comparison of cytolytic and proliferative activities of human and T-cells from peripheral blood against various human tumor cell lines. *Journal of the National Cancer Institute* 83:1564, 1994.

14. Orsini DL, VanGils M, Kooy YMC, Struyk L, Klein G, van den Elsen P, Koning F. Functional and molecular characterization of B-cell-responsive V 1<sup>+</sup> T-cells. *European Journal of Immunology* 24:3199, 1994.
15. Hoffman, T., Theobald, M., Bunjes, D., Weiss, M., Heimpel, H., Heidt, W. Frequency of bone marrow T cells responding to HLA-identical non-leukemic and leukemic stimulator cells. *Bone Marrow Transplantation* 1993; 12: 1.
16. Fisch, P., Malkovska, M., Braakman, E., Sturm, E., Bolhuis, R.L., Prieve, A., Sosman, J.A., Lam, V.A., Sondel, P.M. Gamma/delta T cell clones and natural killer cell clones mediate distinct patterns of non-major histocompatibility-restricted cytotoxicity. *J. Exp. Med.* 1990; 171: 1567.
17. Kaur, I., Voss, S.D., Gupta, R.S., Schell, K., Fisch, P., Sondel, P.M. Human peripheral gamma/delta T cells recognize hsp60 molecules on Daudi Burkitt's lymphoma cells. *J. Immunol.* 1993; 150: 2046.
18. Battistini, L., Salvetti, M., Falcone, B., Raine, C.S., Brosnan, C.F. Gamma delta T cell receptor analysis supports a role for HSP 70 selection of lymphocytes in multiple sclerosis lesions. *Mol. Med.* 1995; 1: 554.
19. Potential antileukemic effect of gamma delta T cells in acute lymphoblastic leukemia. *Leukemia* 1995; 9: 863.
20. Lamb LS, Henslee-Downey PJ, Parrish RS, Godder KT, Thompson J, Lee C, Gee AP. Increased frequency of TCR-<sup>+</sup> T-cells in disease-free survivors following T-cell depleted partially mismatched bone marrow transplantation for leukemia. *Journal of Hematotherapy* 5:503, 1996.
21. Lamb, L.S., Gee, A., Hazlett, L, Musk, P., Parrish, R.S., O'Hanlon, T.P., Geier, S., Folk, R.S., Harris, W.G., McPherson, K., Lee, C., Henslee-Downey, P.J. Influence of T cell depletion method on circulating  $\gamma\delta$  T cell reconstitution and potential role in the graft-versus-leukemia effect *Cytotherapy* (in press).
22. Anasetti, C., Amos, D., Beatty, P.G., Appelbaum, F.R. et al. Effect of HLA compatibility on engraftment of bone marrow transplants in patients with leukemia or lymphoma. *New Engl. J. Med.* 1989; 320: 197.



23. Anasetti, C., Beatty, P.G., Storb, R., Martin, P.J. et al. Effect of HLA incompatibility on graft-versus-host disease, relapse and survival after marrow transplantation for patients with leukemia or lymphoma. *Human Immunol.* 1990; 29: 79.
- 5 24. Beatty, P.G., Clift, R.A., Mickelson, E.M., Nisperos, B.B., Flournoy, N., Martin, P.J., Sanders, J.E., Stewart, P., Buckner, C.D., Storb, R., Thomas, E.D., Hansen, J. Marrow transplantation from related donors other than HLA-identical siblings. *New Engl. J. Med.* 1985; 313: 765.
- 10 25. Gajewski, J., Ceka, M., Champlin, R. Bone marrow transplantation utilizing HLA-matched unrelated donors. *Blood Reviews* 1990; 4: 132.
26. Frame, J.N., Collins, N.H., Cartagena, T., Waldmann, H., O'Reilly, R., Dupont, B. & Kernan, N.A. T cell depletion of human bone marrow: comparison of Campath I plus complement, anti-T cell  $\alpha$ -chain immunotoxin, and soybean agglutinin alone or in combination with sheep erythrocytes or
- 15 immunomagnetic beads. *Transplantation*, 1989; 47: 984.
27. Kernan, N.A., Bordignon, C., Heller, G., Cunningham, I. et al. Graft failure after T cell depleted human leukocyte antigen identical marrow transplants for leukemia. *Blood* 1989; 74: 2227.
28. Martin, P.J., Hansen, J.A., Torok-Storb, B., Durnam, D. et al: Graft
- 20 failure in patients receiving T cell depleted HLA-identical allogeneic marrow transplants. *Bone Marrow Transplantation* 1988; 3: 445.
29. Korngold, R. and Sprent, J. T cell subsets and graft-versus-host disease. *Transpln Proc.* 44, 335, 1987.
30. Ash, R.C., Horowitz, M.M., Gale, R.P., van Bekkum, J.T. et al. Bone
- 25 marrow transplantation from related donors other than HLA-identical siblings: effect of T cell depletion. *Bone Marrow Transplantation* 7, 441, 1991.
31. Henslee-Downey, P.J., Parrish, R., MacDonald, J.S., Romond, E.H., Marciniack, E., Coffey, C., Ciocchi, G., Thompson, Jr. Combined in vitro and in vivo T lymphocyte depletion for the control of graft-versus-host disease following
- 30 haploidentical marrow transplant. *Transplantation* 1886; 61; 738.
32. Lee, C., Henslee-Downey, P.J., Brouillette, M., Pati, A.R., Godder, K., Abhyankar, S.H., Gee, A.P. Comparison of OKT-3 and T10B9 for ex vivo T cell

depletion of partially mismatched related donor bone marrow transplantation.  
*Blood* 1996; 86: 625a.

33. Goldman, J.M., Gale, R.P., Horowitz, M.M., Biggs, J.C., Champlin, R.E., Gluckman, E., Hoffman, R.G., Jacobsen, S.J., Marmot, A., M., McGlave, P.B., Messner, H.A., Rimm, A., A., Rozman, C., Speck, B., Tura, S., Weinger, R.S. & Bortin, M.M. Bone marrow transplantation for chronic myelogenous leukemia in chronic phase: Increased risk for relapse associated with T-cell depletion. *Ann Int Med*, 1988; 108; 806.
34. Hesner, M.J., Endean, D., Caspter, J.T., Horowitz, M.M., Keverer-Taylor, C.A., Roth, M., Flomenberg, N., Drobyski, W.R. Use of unrelated marrow grafts compensates for reduced graft-versus-leukemia reactivity after T cell depleted allogeneic marrow transplantation for chronic myelogenous leukemia *Blood* 1995; 86: 3987.
35. Kolb, H.J., Schattenberg, A., Goldman, J.M., Hertenstein, B., Jacobsen, N., Arcese, W., Ljungman, P., Ferrant, A., Verdonck, L., Neiderweiser, D., van Rhee, F., Mittermueller, J., de Witte, T., Holler, E., Ansari, H. Graft-versus-leukemia effect of donor lymphocyte transfusions in marrow grafted patients. *Blood* 1995; 86: 2041.
36. Slavin, S., Naparstek, E., Nagler, A., Ackerstein, A., Kapelushnik, J., Or, R. Allogeneic cell therapy for relapsed leukemia after bone marrow transplantation with donor peripheral blood lymphocytes. *Exp. Hematol.* 1995; 23: 1553.
37. Antin, J.H. Graft-versus-leukemia: no longer an epiphenomenon. *Blood* 1993; 82: 2273.
38. Barrett, A.J. Strategies to enhance the graft-versus-malignancy effect in allogeneic transplants. *Ann NY Acad Sci* 1996; XX: 203.
39. Truitt, R.L., Johnson, B.D. Principles of graft-versus-leukemia reactivity. *Biol of Blood and Marrow Transplantation* 1995; 1: 61.
40. Datta, A.R., Barrett, A.J., Jiang, Y.Z., Guimaraes, A., Mavroudis, D.A., van Rhee, F., Gordon, A.A., Madrigal, A. Distinct T cell populations distinguish chronic myeloid leukemia cells from lymphocytes in the same

individual: a model for separating GVHD from GVL reactions. *Bone Marrow Transplantation* 1994; 14: 517.

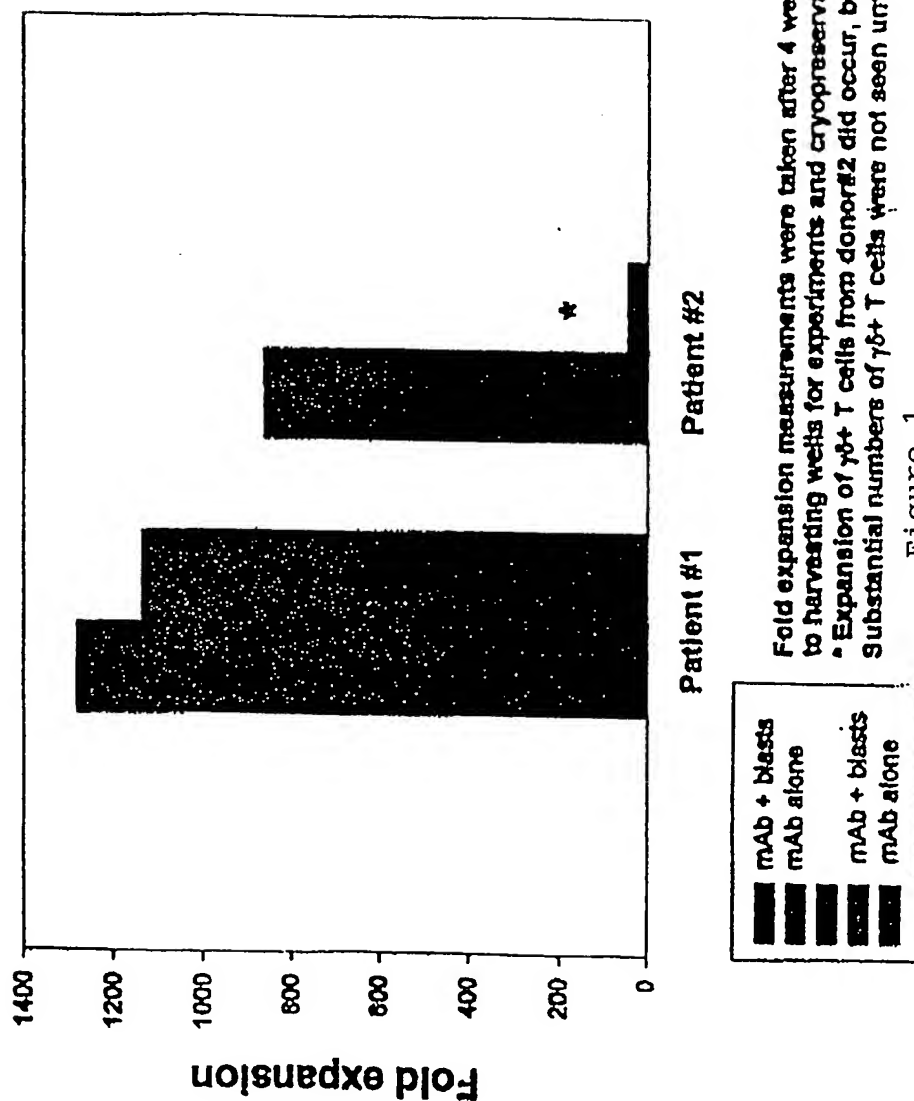
41. Champlin, R. Separation of graft-versus-host disease and graft-versus-leukemia effect against chronic myelogenous leukemia. *Exp. Hematol.* 1995; 23: 1148.
42. Raulet, D.H. The structure, function, and molecular genetics of the  $\gamma/\delta$  cell receptor. *Ann. Rev. Immunol.*, 1989; 7: 175.
43. Bigby, M., Markowitz, J.M., Bleicher, P.A., Grusby, M.J., Simha, S., Siebrecht, M., Wagner, M., Nagler-Anderson, C., Glimcher, L.H. most  $\gamma\delta$  T cells develop normally in the absence of MHC class II antigens. *J. Immunol.* 1993; 151: 4465.
44. Lanier, L. Unusual lymphocytes -  $\gamma\delta$  T cells and NK cells. *The Immunologist* 1995; 3; 182.
45. Schild, H., Mavaddat, N., Litzenberger, C., Ehrlich, E.W., Davis, M.M., Bluestone, J.A., Matis, L., Draper, R.K. Chien, y. The nature of major histocompatibility complex recognition by  $\gamma\delta$ + T cells. *Cell* 1994; 76: 29.
46. Sperling, A.I., Linsley, P.S., Barrett, T.A., Bluestone, J.A. CD-28 mediated costimulation is necessary for the activation of T cell receptor- $\gamma\delta$ + T lymphocytes. *J. Immunol.* 1993; 151: 6043.
47. Wesselborg, S., Janssen, O., Pechhold, K., Kabelitz, D. Selective activation of  $\gamma/\delta$ + T cells by single anti-CD2 antibodies. *J. Exp. Med.* 1991; 173: 297.
48. Avdalovic, M., Fong, D., Formby, B. Adhesion and costimulation of proliferative responses of human  $\gamma\delta$  T cells by interaction of VLA-4 and VLA-5 with fibronectin. *Immunol. Lett.* 1993; 35: 101.
49. Lin, T., Matsuzaki, G., Umesue, M., Omoto, K., Yoshida, H., Harada, M., Singaram, C., Hiromatsu, K., Nomoto, K. Development of  $\gamma\delta$  CD4<sup>-</sup>CD8<sup>+</sup>  $\alpha\alpha$  but not TCR- $\alpha\beta$  CD4<sup>-</sup>CD8<sup>+</sup> $\alpha\alpha$  i-IEL is resistant to cyclosporin A. *J. Immunol.* 1995; 155: 4224.
50. Ellison, C.A., MacDonald, G.C., Rector, E.S., Gartner, J.G.  $\gamma\delta$  T cells in the pathobiology of murine acute graft-versus-host disease. *J. Immunol.* 1995; 155: 4189.

51. Blazar, B.R., Taylor, P.A. Bluestone, J.A., Valleria, D.A. Murine  $\gamma\delta$ -expressing T cells affect alloengraftment via the reconognition of nonclassical major histocompatibility complex class Ib antigens. *Blood* 1996; 87: 4463.
52. Chouaib, F., Porto, P., Delorme, D., Hercend, T. Further evidence  
5 for a g/d T cell receptor-mediated TCT.1/CD48 recognition. *J. Immunol.* 147: 2864; 1991.
53. Hacker, G., Kromer, S., Falk, M., Heeg, K., Wagner, H., Pfeffer, K.  $V\delta 1+$  subset of human  $\gamma\delta$  T cells responds to ligands expressed by EBV-infected Burkitt lymphoma cells and transformed B lymphocytes. *J. Immunol.* 149: 3984;  
10 1992.
54. Marx, S., Wesch, D., Kabelitz, D. Activation of human  $\gamma\delta$  T cells by *Mycobacterium tuberculosis* and Daudi lymphoma cells. *J. Immunol.* 158: 2842; 1997.
55. Chomczynski P, Sacchi N. Single-step method of RNA isolation by  
15 guanidinium thiocyanate-phenol-chloroform extraction. *Anal. Biochem.* 1987; 162:156.
56. O'Hanlon, T.P., Messersmith, W., Dalakas, M.C., Plotz, P.H., and Miller, F.W. T Cell Receptor Gene Expression by Muscle-Infiltrating Lymphocytes in Idiopathic Inflammatory Myopathies. *Clin. Exp. Immunol.* 1995;  
20 100: 519.

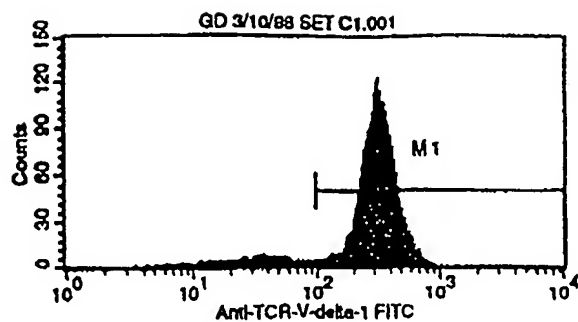
**IN THE CLAIMS:**

1. A cell line comprising substantially purified  $\gamma\delta$ + T lymphocytes.
2. The cell line of claim 1, wherein said  $\gamma\delta$ + T lymphocytes preferentially express the V $\delta$ 1 phenotype.
- 5        3. The cell line of claim 1, wherein said  $\gamma\delta$ + T lymphocytes preferentially express the V $\delta$ 2 phenotype.
4. A method of treating leukemia comprising the steps of:  
administering to a leukemic patient a suitable amount  
of substantially purified  $\gamma\delta$ + T cells; and  
10        monitoring said patient for the presence of leukemia.
5. The method in accordance with claim 4, wherein said  $\gamma\delta$ + T lymphocytes preferentially express the V $\delta$ 1 phenotype.
6. The method in accordance with claim 4, wherein said  $\gamma\delta$ + T lymphocytes preferentially express the V $\delta$ 2 phenotype.
- 15        7. A method of preparing substantially purified  $\gamma\delta$ + T cells comprising the steps of:  
incubating donor  $\gamma\delta$ + T cells with irradiated recipient leukemia  
to allow activation and expansion of  $\gamma\delta$ + T cells; and  
harvesting said  $\gamma\delta$ + T cells.

# EXPANSION OF DONOR $\gamma\delta$ + T CELLS IN CULTURE

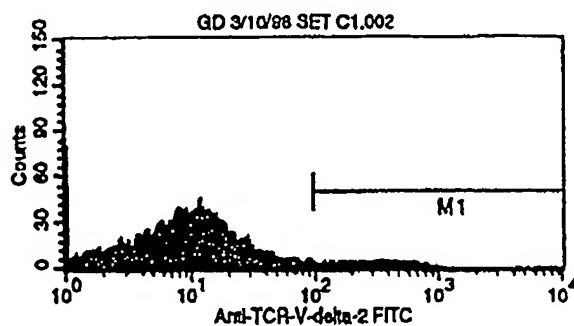


2/9

 $\gamma\delta$ + T cells cultured on pan- $\delta$  MAb with blasts.

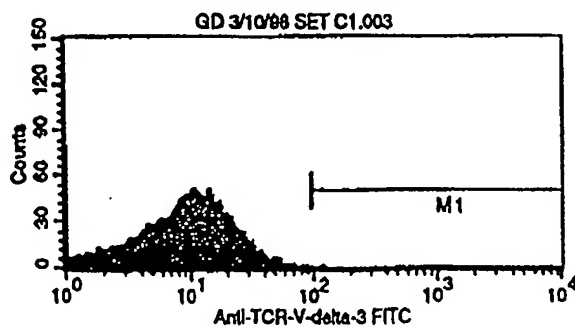
File: GD 3/10/98 SET C1.001  
 Acquisition Date: 10-Mar-98  
 Gate: G3

Marker	Left, Right	Events	% Gated	% Total	CV
All	1, 9910	9658	100.00	96.58	48.
M1	97, 9910	8613	89.18	86.13	33.



File: GD 3/10/98 SET C1.002  
 Acquisition Date: 10-Mar-98  
 Gate: G3

Marker	Left, Right	Events	% Gated	% Total	CV
All	1, 9910	9622	100.00	96.22	281.
M1	97, 9910	658	6.84	6.58	60.

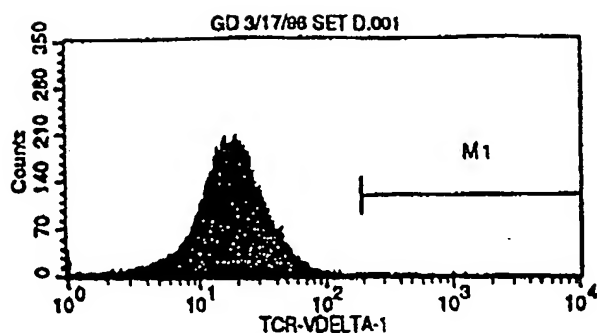


File: GD 3/10/98 SET C1.003  
 Acquisition Date: 10-Mar-98  
 Gate: G3

Marker	Left, Right	Events	% Gated	% Total	CV
All	1, 9910	8598	100.00	95.98	87.4
M1	97, 9910	7	0.07	0.07	21.4

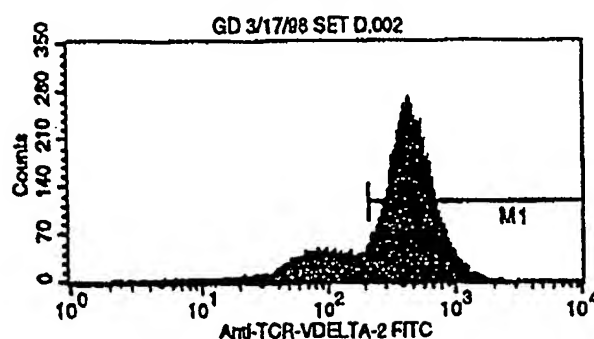
Figure 2

3/9

 $\gamma\delta$ + T cells cultured on pan- $\delta$  MAb without blasts.

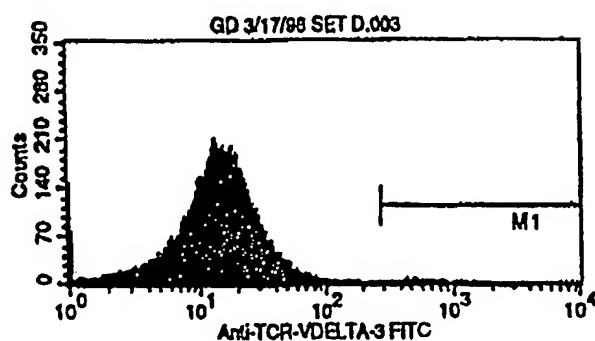
File: GD 3/17/98 SET D.001  
 Acquisition Date: 18-Mar-98  
 Gate: G3

Marker	Left, Right	Events	% Gated	% Total	CV
All	1, 9910	29133	100.00	97.11	91
M1	195, 9910	34	0.12	0.11	62



File: GD 3/17/98 SET D.002  
 Acquisition Date: 18-Mar-98  
 Gate: G3

Marker	Left, Right	Events	% Gated	% Total	CV
All	1, 9910	29252	100.00	97.51	57
M1	219, 9910	23363	79.83	77.84	36



File: GD 3/17/98 SET D.003  
 Acquisition Date: 18-Mar-98  
 Gate: G3

Marker	Left, Right	Events	% Gated	% Total	CV
All	1, 9910	29231	100.00	97.44	185.
M1	288, 9910	108	0.37	0.36	37.

Figure 3



4/9

# $\gamma\delta$ T CELL PHENOTYPE - PATIENT #1

CD4+/CD8+ DEPLETED MNC CULTURED FOR 6 WEEKS ON  
PAN- $\delta$  mAB + RECIPIENT BLASTS

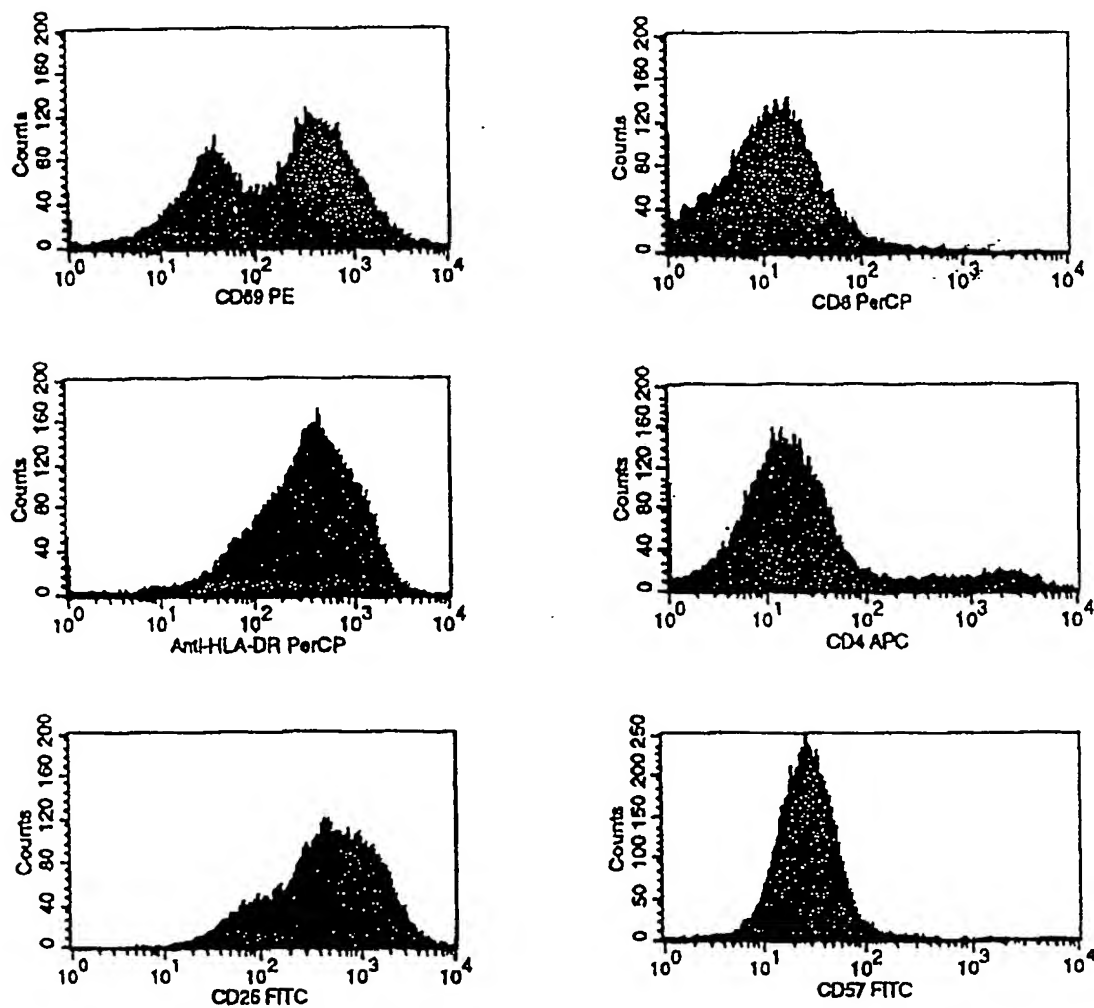
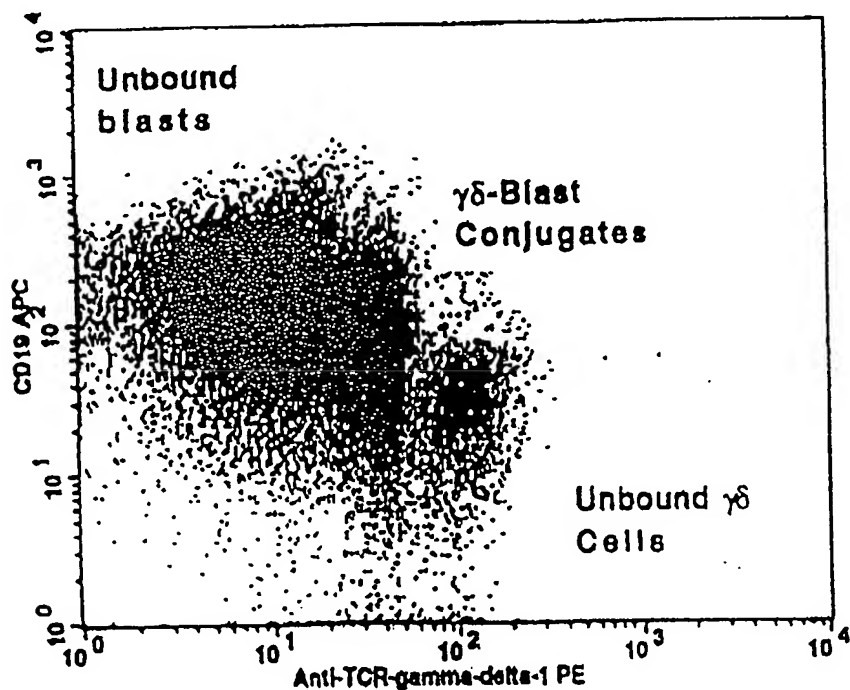


Figure 4

5/9

# FLOW CYTOMETRIC BINDING ASSAY SHOWING ACTIVATED DONOR $\gamma\delta$ + T CELLS BINDING TO RECIPIENT LEUKEMIC CD19+ BLASTS

PATIENT #1



PATIENT #2

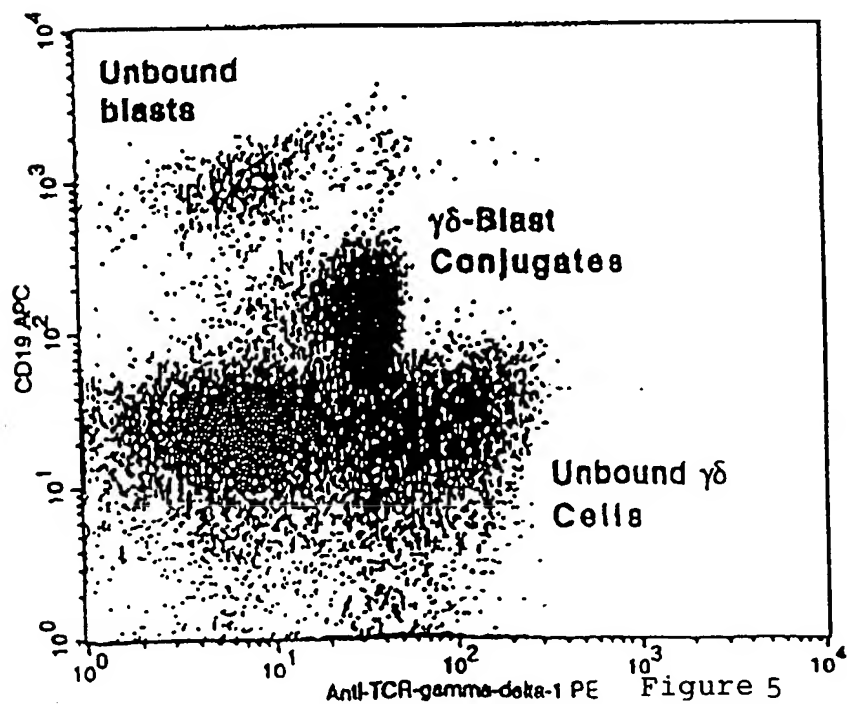
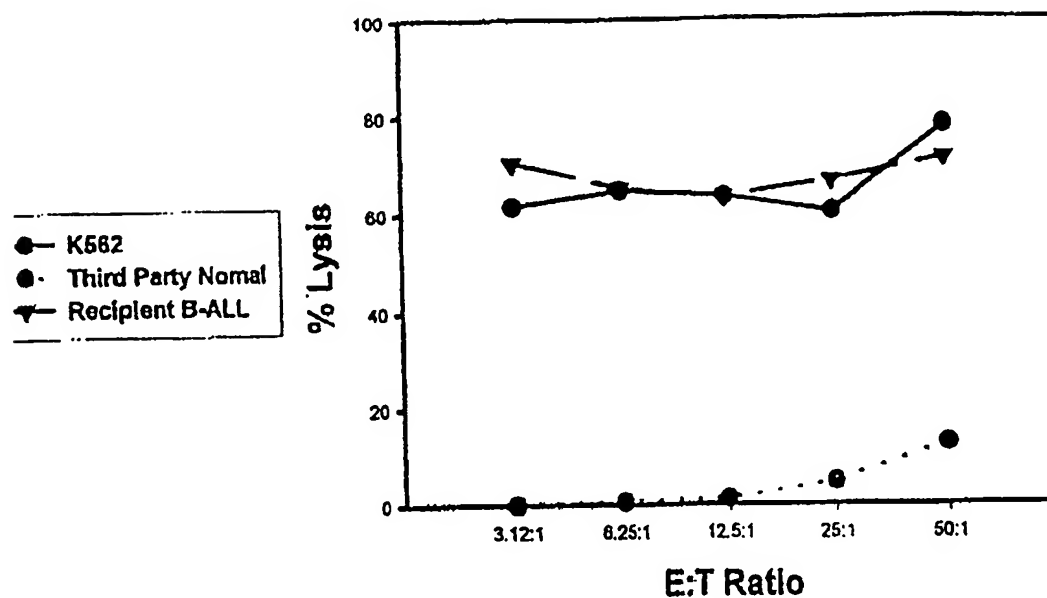


Figure 5

6/9

CYTOTOXICITY OF DONOR  $\gamma\delta$ + T CELLS

## Patient #1



## Patient #2

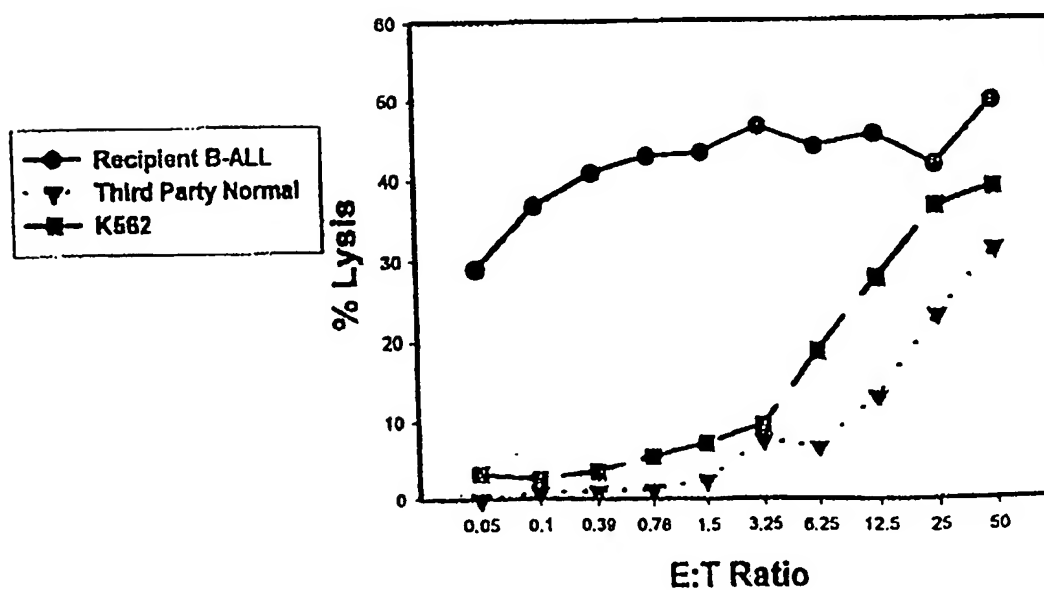


Figure 6

7/9

# CYTOTOXICITY OF EXPANDED $\gamma\delta$ + T CELLS AGAINST VARIOUS CELL LINES

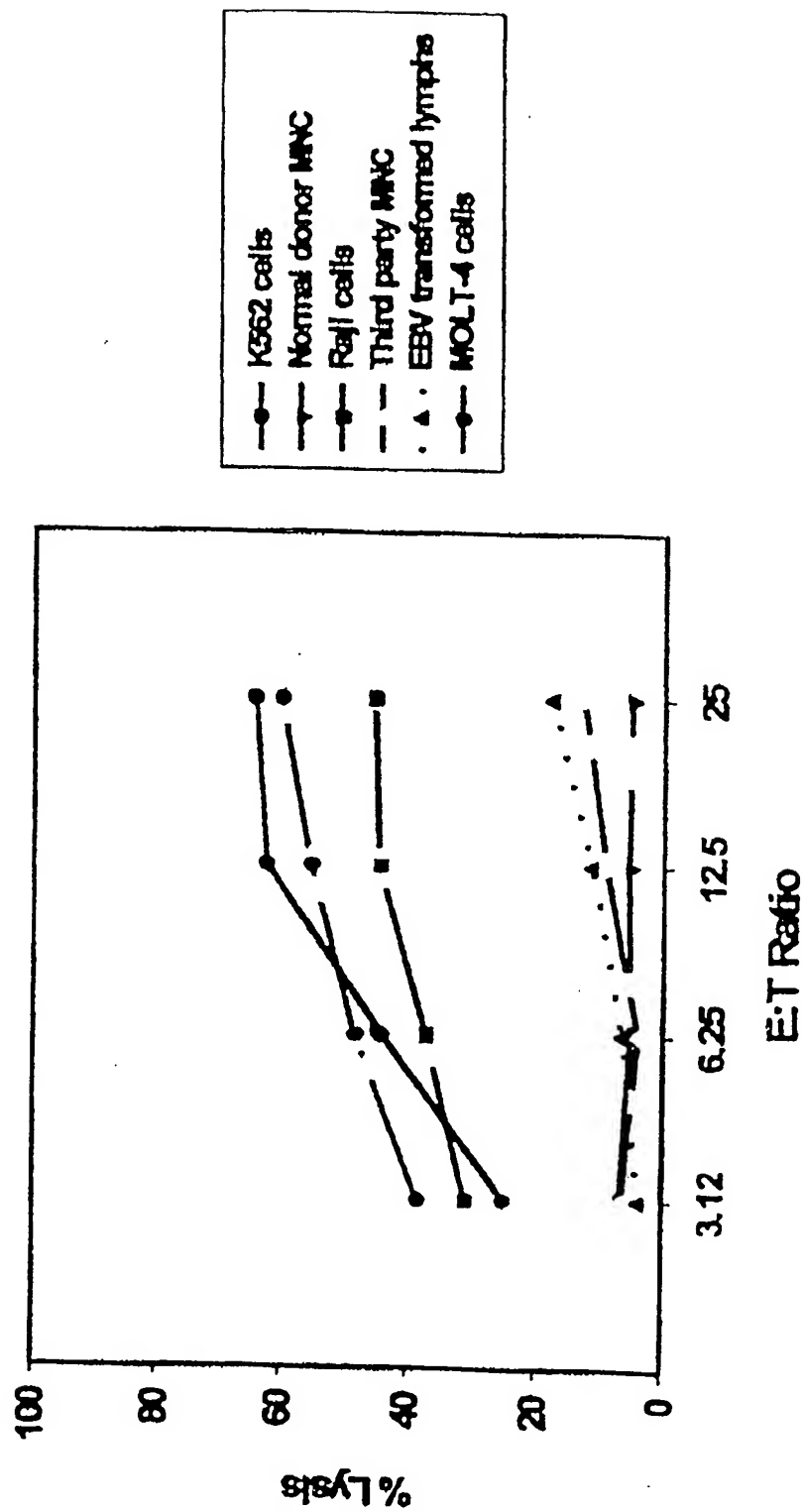


Figure 7

8/9

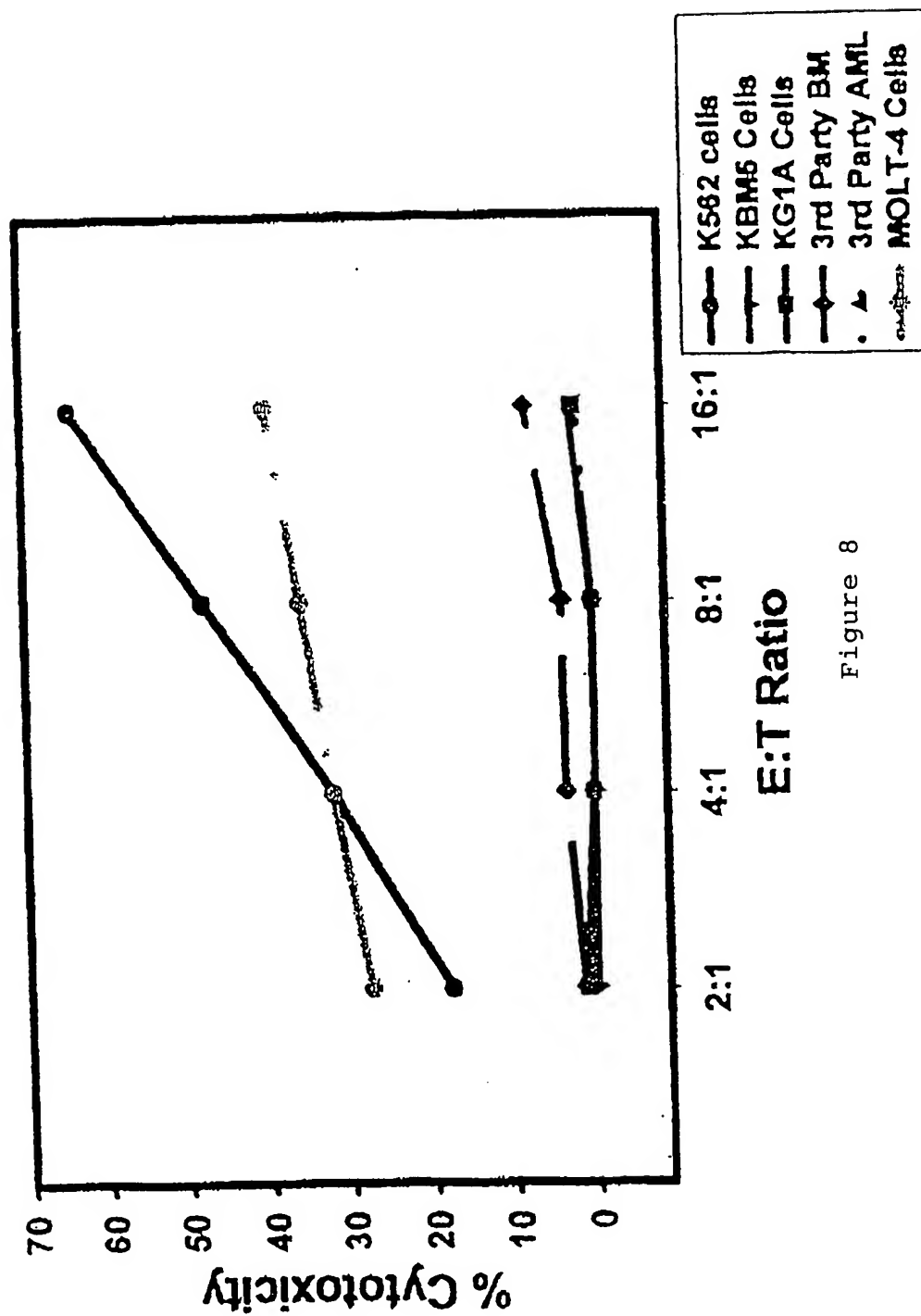


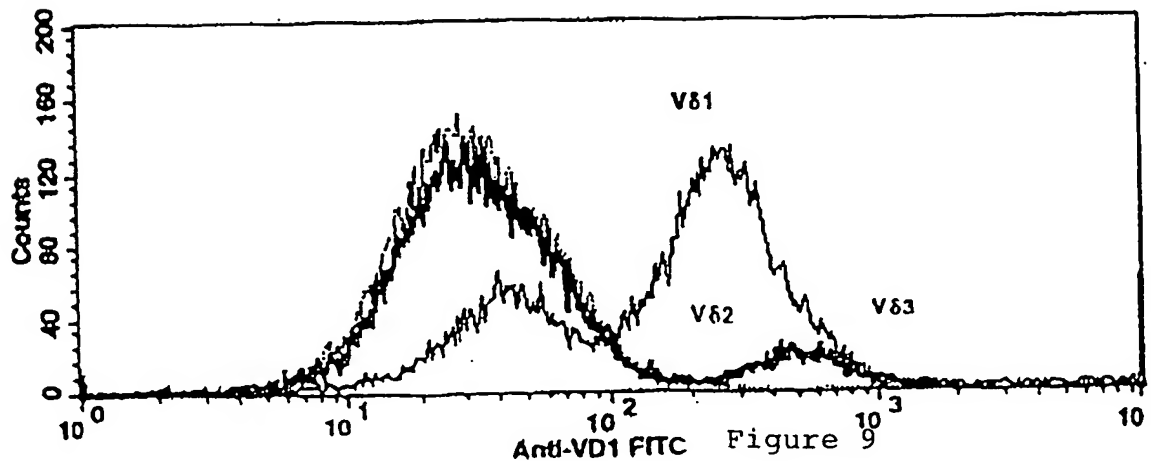
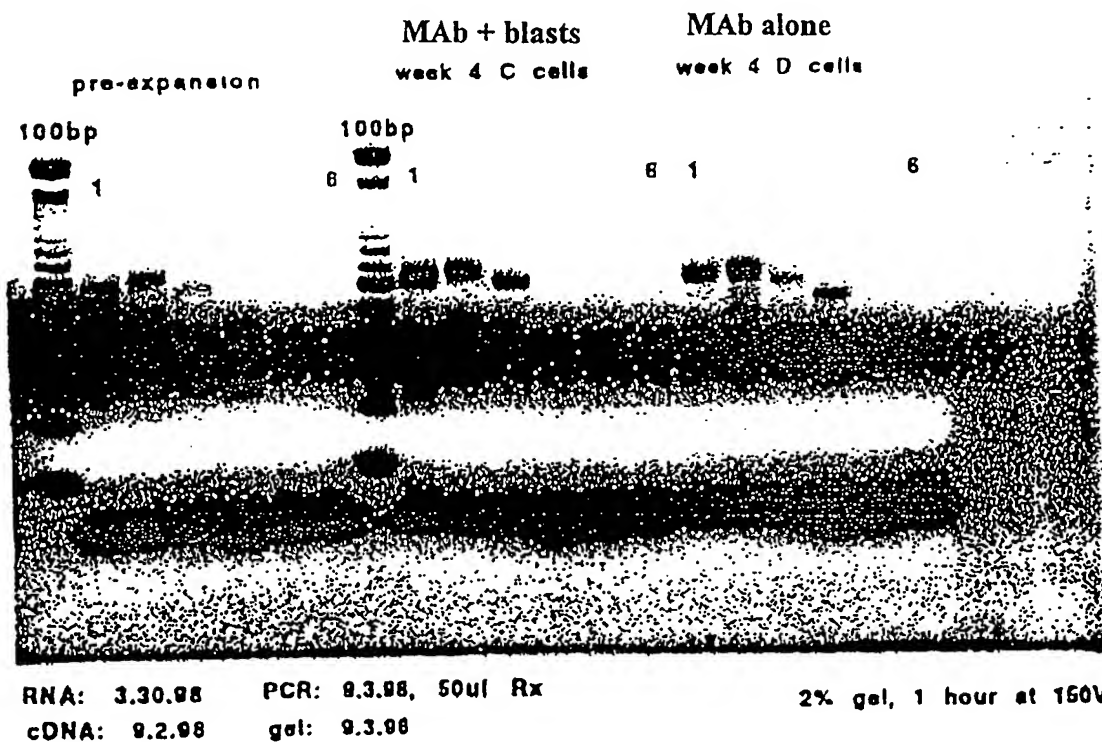
Figure 8

9/9

## mRNA AND SURFACE EXPRESSION OF V $\delta$ SUBTYPES - PATIENT #1

CD4+/CD8+ DEPLETED DONOR MNC CULTURED FOR 4 WEEKS ON PAN- $\delta$  mAB + RECIPIENT BLASTS

A8 versus SL expansion #2



## INTERNATIONAL SEARCH REPORT

International application No.

PCT/US00/01867

## A. CLASSIFICATION OF SUBJECT MATTER

IPC(7) : C12N 15/07, 15/01, 5/06

US CL : 435/375, 355, 361, 372

According to International Patent Classification (IPC) or to both national classification and IPC

## B. FIELDS SEARCHED

Minimum documentation searched (classification system followed by classification symbols)

U.S. : 435/375, 355, 361, 372

Documentation searched other than minimum documentation to the extent that such documents are included in the fields searched

Electronic data base consulted during the international search (name of data base and, where practicable, search terms used)

JPO, EPO, DERWENT, MEDLINE, BIOSIS, CAPLUS

search terms: lymphocytes, gamma delta, leukemia

## C. DOCUMENTS CONSIDERED TO BE RELEVANT

Category*	Citation of document, with indication, where appropriate, of the relevant passages	Relevant to claim No.
X, Y	EP 0 786,664 A1 (ROMAGNE et al.) 30 July 1997, see entire document, especially page 2, lines 25-30.	X 1,4,7 --- Y 2-3,5-6
A	WO 98/33891 A (BELL et al.) 06 August 1998.	1-7

☐ Further documents are listed in the continuation of Box C. ☐ See patent family annex.

* Special categories of cited documents:	*T* later document published after the international filing date or priority date and not in conflict with the application but cited to understand the principle or theory underlying the invention
*A* document defining the general state of the art which is not considered to be of particular relevance	*X* document of particular relevance; the claimed invention cannot be considered novel or cannot be considered to involve an inventive step when the document is taken alone
*E* earlier document published on or after the international filing date	*Y* document of particular relevance; the claimed invention cannot be considered to involve an inventive step when the document is combined with one or more other such documents, such combination being obvious to a person skilled in the art
*L* document which may throw doubts on priority claim(s) or which is cited to establish the publication date of another citation or other special reason (as specified)	*A* document member of the same patent family
*O* document referring to an oral disclosure, use, exhibition or other means	
*P* document published prior to the international filing date but later than the priority date claimed	

Date of the actual completion of the international search 05 MAY 2000	Date of mailing of the international search report 07 JUN 2000
Name and mailing address of the ISA/US Commissioner of Patents and Trademarks Box PCT Washington, D.C. 20231 Facsimile No. (703) 305-4242	Authorized officer <i>Jan Patten</i> PATRICIA PATTEN Telephone No. (703) 308-1189